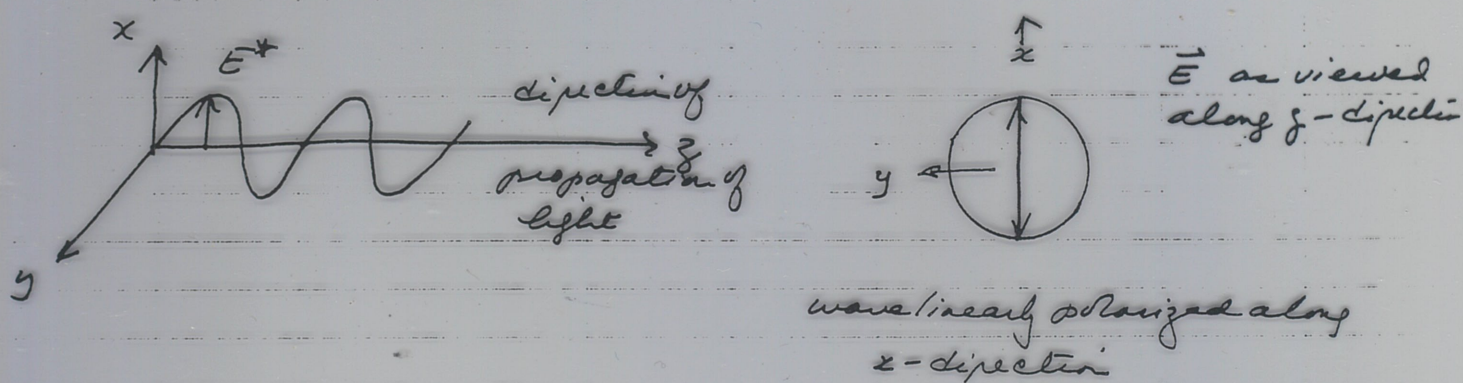


Lecture 18
Physical Chemistry
for Life Scientists
National Tsing-Hua
University
June 2, 2001

①

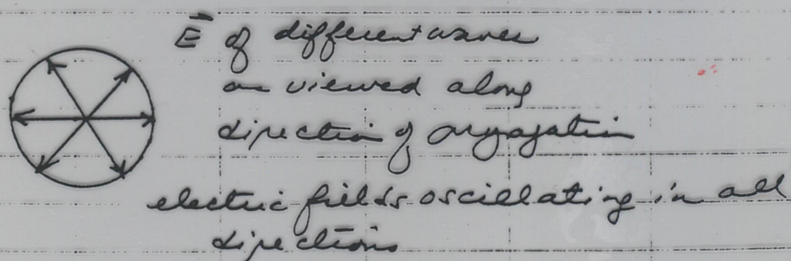
Optical Rotation, Optical Rotatory Dispersion, Circular Dichroism (Interaction of molecules with circularly polarized light)

→ Polarization of Light



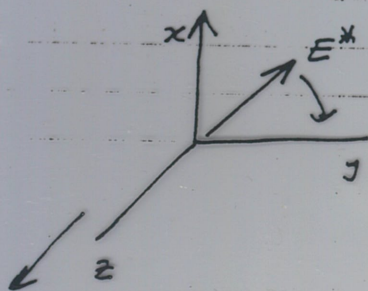
If all the waves have $\vec{E}$ linearly polarized as above, Light is plane-polarized (plane defined by electric field direction and direction of propagation of light wave)

Unpolarized Light



Plane-polarized light can be produced by passing unpolarized light thru a highly anisotropic material (e.g. Polaroid), which absorbs all light with $\vec{E}$ pointing in one-direction (e.g. Horizontal), thereby leaving only rays whose electric field point vertically. Another method is to pass the unpolarized light through a Nicol prism, which removes light polarized in one-direction by reflection.

Plane-polarized light may be considered as a superposition of left and right-circularly polarized light in equal proportions



$$E_R = \vec{i}E_0^* \cos \omega t + \vec{j}E_0^* \sin \omega t \quad \text{clockwise}$$

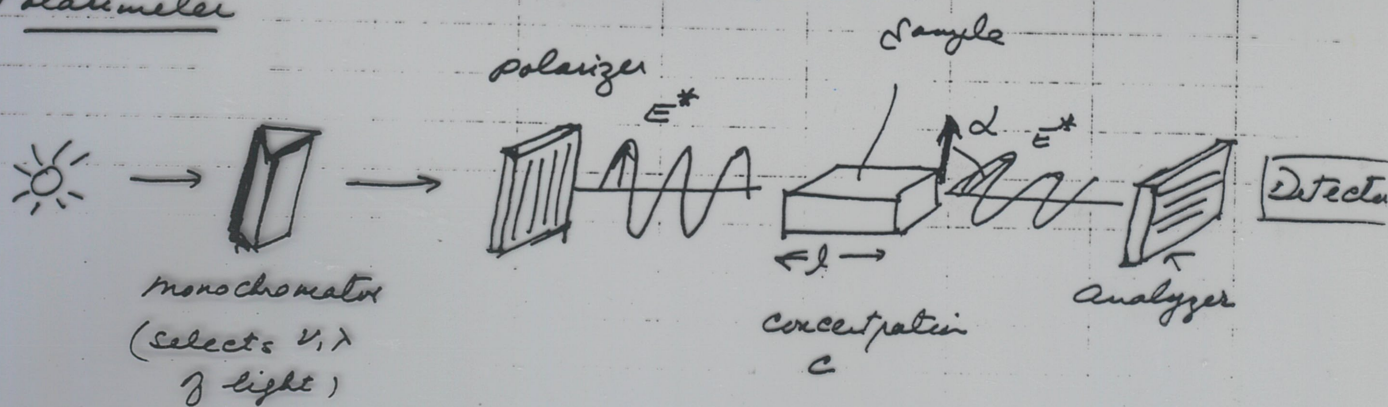
$$E_L = \vec{i}E_0^* \cos \omega t - \vec{j}E_0^* \sin \omega t \quad \text{counter-clockwise}$$

$$E_R + E_L = \vec{i}2E_0^* \cos \omega t$$

as on coming
light beam
is viewed.

→ Optical Rotation

When plane-polarized light passes thru an optically active substance, the plane of polarization of the light is rotated.

Polarimeter

$\alpha \equiv$ angle of rotation of the plane of polarization

Detector measures intensity of transmitted light.

Null method is usually used, i.e., analyzer is set such that light that is passed by the first polarizer

(say vertically polarized) is absorbed by the analyzer.

If the sample rotates the plane of the ~~plane~~ polarization of the light incident on it, say by angle α , then the analyzer must be rotated by angle α to attain null at detector, i.e., polarizer and analyzer are rotated by $90^\circ + \alpha$ w.r.t. each other.

Specific rotation (in degrees) is defined as follows:

$$[\alpha]_{\lambda}^t = \frac{100\alpha}{l c}$$

l = path length of cell in decimeters (1 decimeter = 10 cm)

λ = wavelength of light

c = concentration of solute in grams per 100 ml solution

t = temp. in $^\circ\text{C}$

Examples

D-D glucose in water

$$[\alpha]_D^{20} = +112.2$$

Na-D-line

20°C

+ plane of polarization rotated to right as one looks at beam.

L-serine in water

$$[\alpha]_D^{20} = -6.8$$

For macromolecules, the optical rotation is usually expressed as the ~~reduced~~ ^{reduced} residue rotation, defined by

$$[m'] = \frac{3}{n^2 + 2} \frac{m}{100} [\alpha]_{\lambda}^t$$

where m = mean residue molecular weight

n = refractive index of the solution

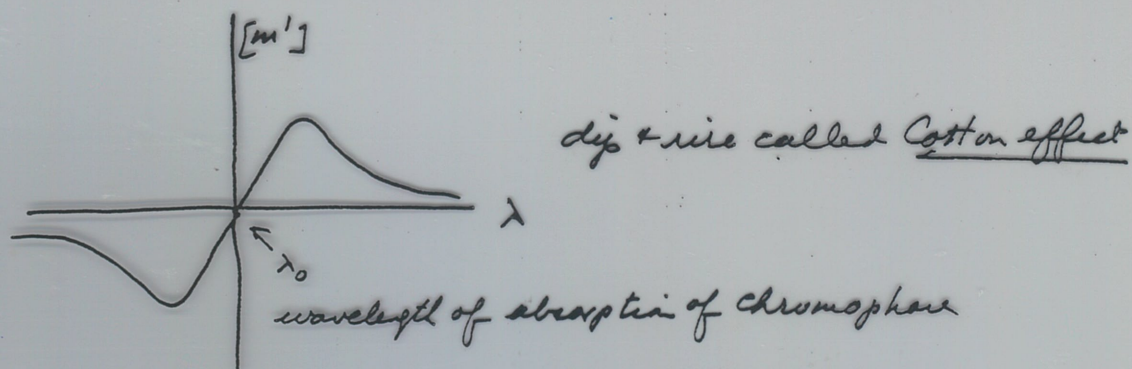
→ Optical Rotatory Dispersion

Optical rotatory dispersion is the dependence of optical rotation on the wavelength of incident light.

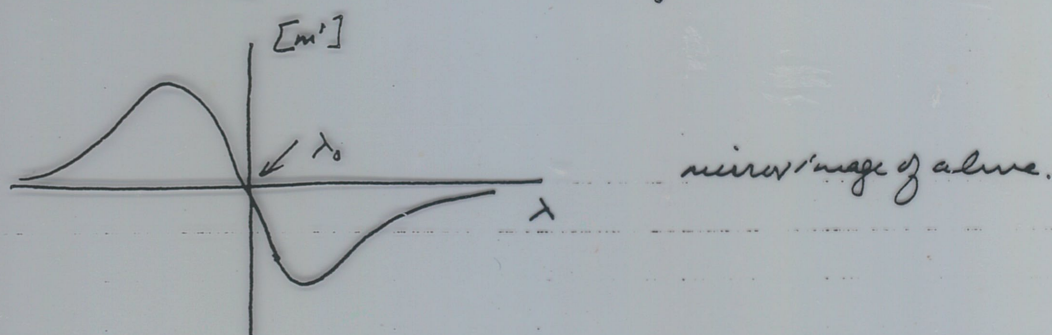
α is measured at each wavelength by a spectropolarimeter and $[m']$ is plotted against λ

Optical Rotation exhibits rise & dip similar to dispersion of the refractive index near chromophore absorptions

ORD of a single asymmetric optically active chromophore



For the enantiomorph (mirror image) of chromophore,



→ Circular Dichroism (CD)

Circular Dichroism is the differential absorption of a sample for left-circularly polarized and right-circularly polarized light.

$$A_L - A_R$$

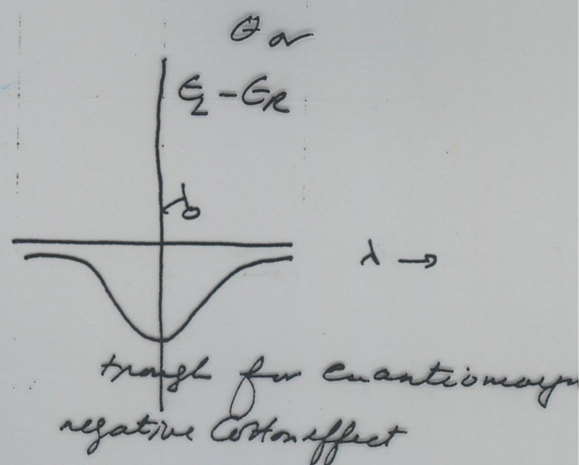
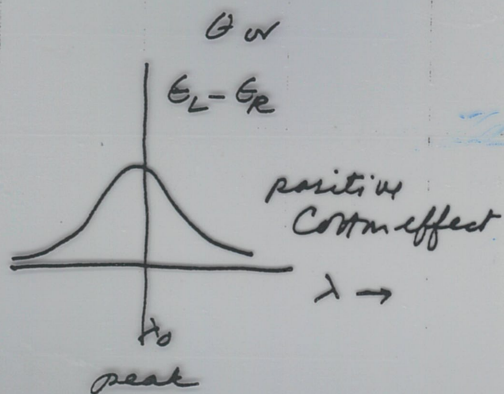
absorbance difference

$$\propto \epsilon_L - \epsilon_R$$

extinction coefficient difference

Recall $\epsilon = A / \text{[]} l c$

CD
for a single
optically active
chromophore

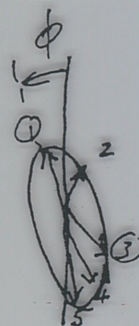
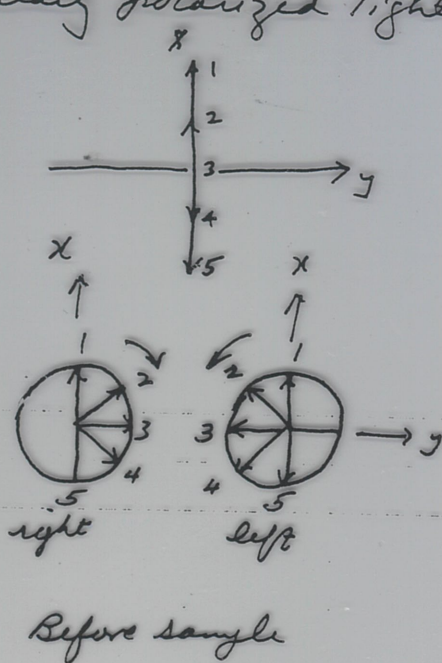


CD or $E_L - E_R$ determined by measuring ellipticity following absorption of linear or plane polarized light or by measuring E_L and E_R using left- and right-circularly polarized light respectively. The latter approach is obvious.

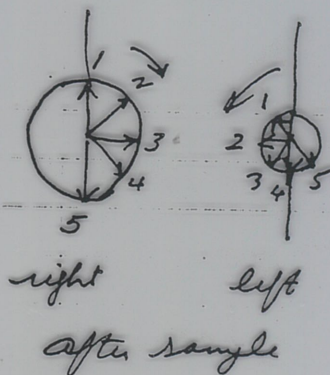
Discuss experiment using plane polarized light.

Recall, can decompose plane polarized light into left- and right circularly polarized light.

1, 2, 3, 4, 5
equal (increasing)
intervals of time



Elliptically
polarized
light produced
by passing
light thru
optically active
sample



left circle
smaller radius
due to preferential
absorption
of left
circularly
polarized
component

Effect of an optically active
sample on two circularly polarized
components

$$\text{Ellipticity} = \tan^{-1} \left(\frac{\text{minor axis}}{\text{major axis}} \right)_{\text{ellipse}} = \theta$$

$$= 2.303 (A_L - A_R) 180 / 4\pi \text{ degree}$$

$$\text{molar ellipticity} = [\theta] = 100 \theta / C \cdot l$$

$$= 2.303 (E_L - E_R) l \cdot c \cdot \frac{100}{C \cdot l} \cdot \frac{180}{4\pi} \text{ degree}$$

$$= 2303 \times \frac{9}{2\pi} (E_L - E_R)$$

$$= 3,300 (E_L - E_R)$$

Circular Birefringence

If the two circularly polarized components are absorbed to different extents at any wavelength, then it follows that the sample will also have a different index of refraction (n) for the two components at virtually all wavelengths. One component will propagate more rapidly than through the other the medium. The result is a phase shift between the two components, proportional to the refractive index difference, $n_L - n_R$. This effect is called circular birefringence. When the two components are combined, the phase shift results in a permanent rotation of the long axis of the elliptically polarized light.

$$\phi = 180 \cdot l \cdot (n_L - n_R) / \lambda \text{ degree}$$

$$* [\phi] = 100 \phi / C \cdot l.$$

ORD and CD are related

$$[\phi(\lambda)] = \frac{2}{\pi} \int_0^{\infty} \frac{[\theta(\lambda')]}{\lambda'^2 - \lambda^2} d\lambda' \quad \text{optical rotation}$$

$$[\theta(\lambda)] = -\frac{2\lambda}{\pi} \int_0^{\infty} \frac{[\phi(\lambda')]}{\lambda'^2 - \lambda^2} d\lambda' \quad \text{circular dichroism}$$

Order of magnitudes

For typical protein or nucleic acid solutions at $10^{-4} M$
Chromophore concentrations,

$$\phi \sim 0.01 \text{ to } 0.1^\circ \text{ for a 1 cm sample}$$

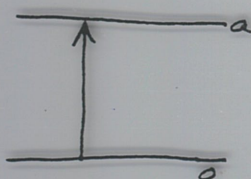
Current instruments can detect rotation as small as 10^{-4} degree.

θ typically 0.03% to 0.3% of total absorption, which can be determined quite accurately with modern instrumentation measuring $E_L - E_R$ by left & right circularly polarized light.

θ , ellipticity is small to measured directly.

Physical origin of CD and ORD

Absorption spectroscopy



Electric dipole transition

Dipole strength

$$= \left| \int \Psi_0^* \vec{\mu} \Psi_a d\tau \cdot \int \Psi_a^* \vec{\mu} \Psi_0 d\tau \right|$$

$$= \left| \int \Psi_0^* \vec{\mu} \Psi_0 d\tau \right|^2$$

Magnetic dipole transition

Dipole strength

$$= \left| \int \Psi_0^* \vec{m} \Psi_a d\tau \right|^2$$

where $\vec{m} = \sum_i \frac{e}{2m_e c} (\vec{r}_i \times \vec{p}_i)$

Circular Dichroism

Rotational Strength

$$= \text{Im} \left(\int \Psi_0^* \vec{\mu} \Psi_a d\tau \cdot \int \Psi_a^* \vec{m} \Psi_0 d\tau \right)$$

↑
dot product

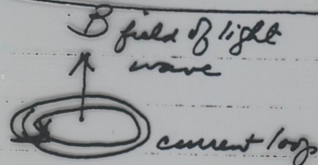
pure electronic absorption



$$\left| \int \Psi_0^* \vec{\mu} \Psi_a d\tau \right|^2$$

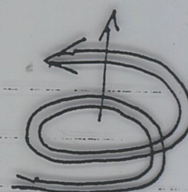
light induced linear displacement

pure magnetic absorption



$$\left| \int \Psi_a^* \vec{m} \Psi_0 d\tau \right|^2$$

light-induced current loop



optical activity

$$\text{Im} \int \Psi_0^* \vec{\mu} \Psi_a d\tau \cdot \int \Psi_a^* \vec{m} \Psi_0 d\tau$$

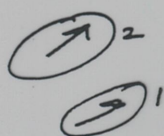
helical structure facilitates flow of charge

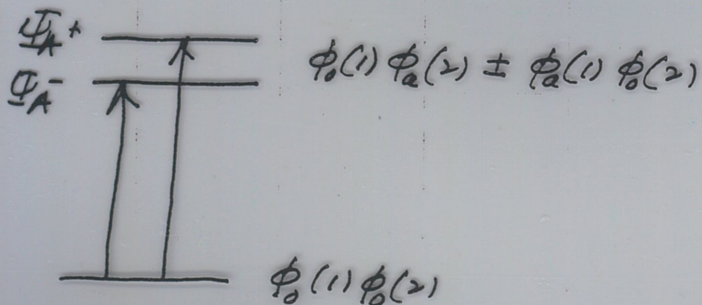
A light wave contains both oscillating electric and magnetic components. The electric component usually dominates pure absorption properties because magnetic effects are small. However, optical activity is a phenomenon that involves combined electric and magnetic interactions. Note dot product — critical feature. This means that for a molecule to be optically active, $\int \Psi_0^* \vec{m} \Psi_0 d\tau$ must have a component parallel to $\int \Psi_0^* \vec{\mu} \Psi_0 d\tau$. The molecule must be asymmetric for this to occur, otherwise there could be no preferred helical direction.

CD in Biomacromolecules

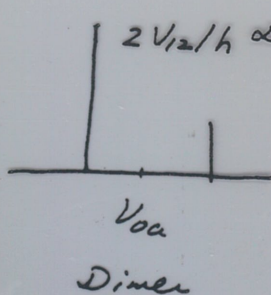
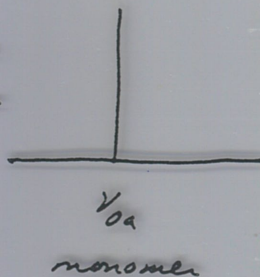
Most important contribution to CD in biomacromolecules arises in change in optical activity when a set of chromophores is assembled into an asymmetric macromolecular structure.

Consider 2 interacting chromophores; assume interaction is electric dipole-dipole interaction

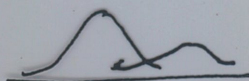




absorption spectrum



electric dipole allowed



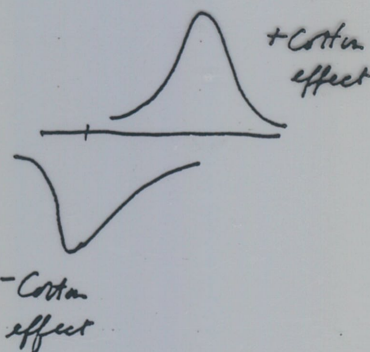
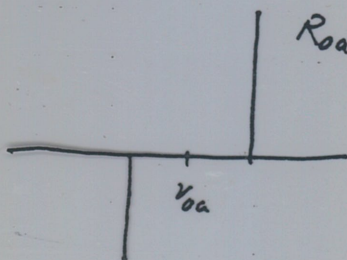
absorption spectrum

if V_{12} small

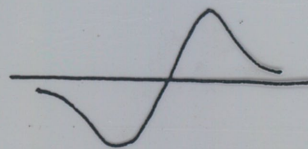


small exciton splitting

CD



Composite for 2 transition



evidence for 2 interacting chromophores.

exciton interaction contribution to CD depends on distance between chromophores ($\propto R_{12}$), & geometry of molecule. Can be large, even if monomeric chromophores are not optically active.

Excitation contribution to $R_{0a} = 0$ if chromophores are coplanar, or if they are oriented parallel or perpendicular to one another.

Example (1) poly L-alanine

a) No contribution from side chain

b) There is near UV $\pi-\pi^*$ and $n-\pi^*$ electronic transition of peptide

CD spectrum on next page.

CD spectrum sensitive to α -helix, β -sheet, random coil

(2) Membrane Proteins

Transmembrane domains are mainly α -helical.

ORD away from λ_{0a}

Optical Rotation at λ far removed from absorption band

$$[m'] = \frac{96N\pi}{hc} \left(\frac{n^2+2}{3} \right) \left(\frac{\lambda_{0a}^2 R_{0a}}{\lambda^2 - \lambda_{0a}^2} \right) \quad \text{for 1 absorption band at } \lambda_{0a}$$

Form of equation explains why molecules such as sucrose can demonstrate substantial optical rotation with visible light even though they cannot absorb this light

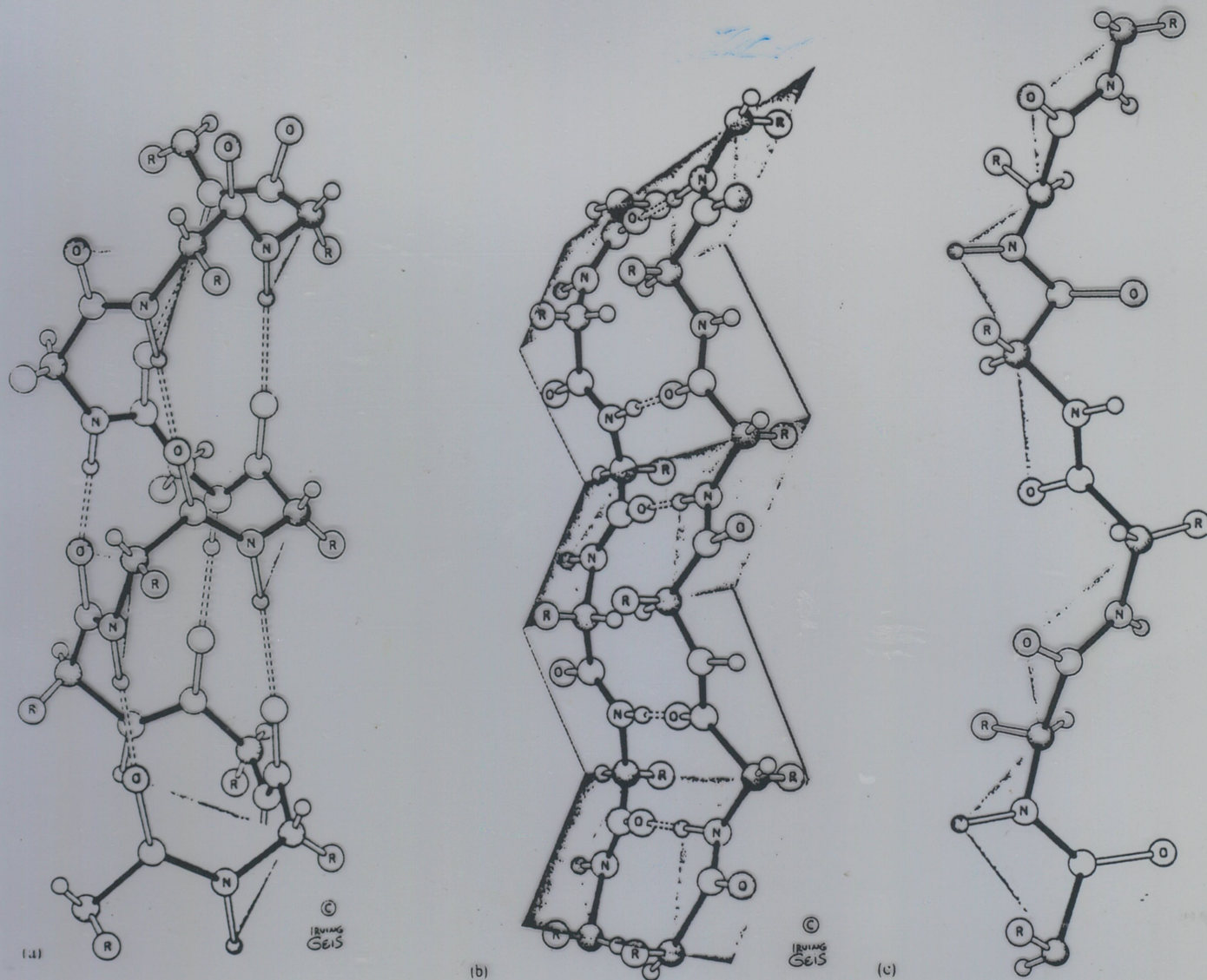


Figure 2-23

Three of the most common polypeptide secondary structures. Only atoms in the peptide-chain backbone are shown. A parallelogram indicates the plane of each peptide bond. (a) The right-handed α helix. (b) Two antiparallel β -sheet strands. Note how alternate side chains will be placed on opposite sides of the sheet. (c) The left-handed polyproline II helix. [Drawings by Irving Geis.]

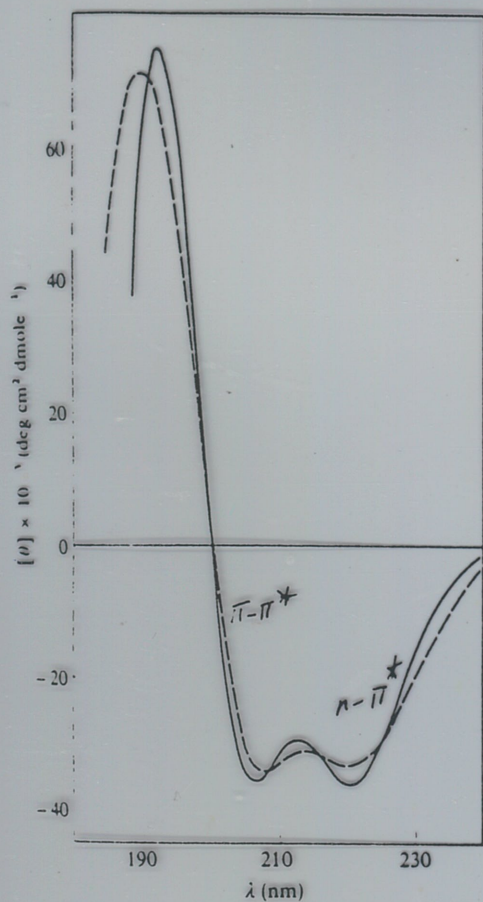


Figure 8-8

CD spectrum of poly-L-alanine in an α -helical conformation. Calculated (dashed line) and observed (solid line) spectra are shown. [After R. W. Woody, *J. Chem. Phys.* 49:4797 (1968).]

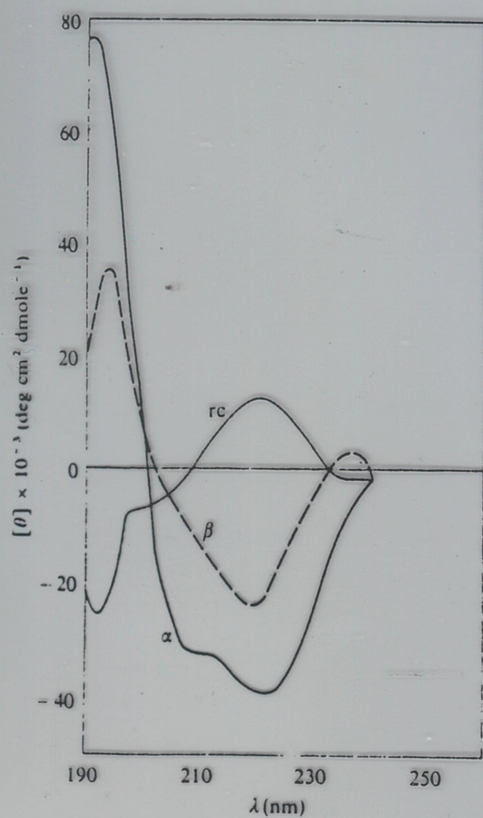


Figure 8-9

CD spectra for α -helix, β -sheet, and random coil conformations, extracted from the spectra of proteins of known three-dimensional structure by the Wetlauffer method. [After V P Saxena and D. B. Wetlauffer, *Proc. Natl. Acad. Sci. USA* 66:969 (1971).]

For more than 1 absorption band,

$$[m'] = \frac{96 N \pi}{h c} \frac{n^2 + 2}{3} \sum_i \frac{\lambda_{0i}^2 R_i}{\lambda^2 - \lambda_{0i}^2}$$